

Centre for AIDS Reagents.



Data Sheet

NAME:	pSHIV-89.6 3'
REPOSITORY REFERENCE:	ARP2123
PROVIDED:	5.78 μg plasmid DNA. Propagate in <i>E. coli</i> XL1-Blue grown at 30°C.
DESCRIPTION:	SHIVs are chimeric simian/human immunodeficiency viruses composed of SIVmac239 modified to include HIV-1 env and the associated auxiliary HIV-1 genes tat, vpu, and rev. The SHIV-89.6 3' construct encodes the HIV-1 portions of the chimeric virus, cloned into the pBluescript II KS(+) vector (Stratagene). The insert size is 5.9 kb, and the plasmid size is 8.9 kb.
CHARACTERISTICS:	The HIV-1 env sequences were derived from the macrophage-tropic isolate HIV-1 89.6. Infectious virus can be generated by ligating pSHIV- 89.6 5' and pSHIV-89.6 3', and transfecting CEMx174 cells with the resulting construct.
STORAGE:	-70°C
SOURCE:	Dr. Joseph Sodroski (Courtesy of the AIDS Research and Reference Reagent Program, Division of AIDS, NIAID, NIH.)

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REFERENCE:

Karlsson GB, Halloran M, Li J, Park IW, Gomila R, Reimann KA, Axthelm MK, Iliff SA, Letvin NL, Sodroski J. Characterization of molecularly cloned simian-human immunodeficiency viruses causing rapid CD4+ lymphocyte depletion in rhesus monkeys. *J Virol* **71**:4218-4225, 1997.

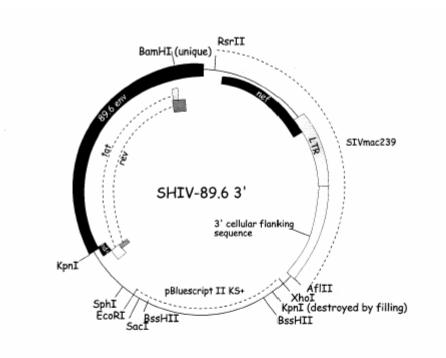
ACKNOWLEDGEMENTS:

Publications should acknowledge the donor of the reagent and the Programme EVA Centre for AIDS Reagents. Suggested wording can be found on our website

http://www.nibsc.ac.uk/spotlight/aidsreagent/index.html in the "Acknowledgements" section.

Please also ensure that you send us a copy of any papers resulting from work using reagents acquired through CFAR (this can be electronically or as a paper copy)

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- 1. EcoRI-XhoI fragment of HXBc2 (5743-8897) cloned into pBluescript II KS+
- 2. RsrII and AfIII sites introduced into HXBc2:

8796 8819
GATGGGTGGCAAGTGGTCAAAAAG
C ACC TT G
RSrII Aflii

3. RsrII site introduced into SIVmac239 nef-open:

9313 9327
TGAACTGACCTACCT
G G
RsrII

- 4. RsrII-AflII fragment of SIVmac239 nef open inserted into the modified HXBc2 subclone.
- 5. SphI site and last two codons of SIV vpr were introduced into HXBc2 sequence:

5808 5822 TACTCGACAGAGGAG GC TGCT T

TRANSFECTION OF CEMX174 CELLS FOR SIV OR SHIV PRODUCTION

DIGESTION

1. Digest 5 μg each proviral half with the appropriate restriction enzymes in a total volume of 80 μl. Remove a 5 μl aliquot and run a gel to make sure digestion has gone to completion.

SHIV-KB9 Cut the 5' half clone with SphI + XhoI
Digest: Cut the 3' half clone with SphI + NotI

SHIV-89.6

Digest: Cut the 5' half clone with SphI + ClaI

Cut the 3' half clone with SphI + AflII

Phenol/chloroform extract the digested DNA once. Precipitate with ethanol using standard procedures.

Resuspend pellets in 20 μ l dH2O and set up ligations in a final volume of 50 μ l, using the total 20 μ l volume of each half. Ligate for at least 3 hours at 17°C.

TRANSFECTION

- 1. Prepare 2M Tris buffer, pH 7.3, and 50 mM Tris buffer, pH 7.3. Filter sterilize.
- 2. Prepare DEAE-dextran at 25 mg/ml in the 50 mM Tris buffer, pH 7.3 (0.25 g DEAE-dextran in 10 ml). Filter sterilize.
- 3. Prepare DME/DEAE by adding 1.25 ml of the 2M Tris buffer, pH 7.3, and 0.25 ml of the 25 mg/ml DEAE-dextran solution into 48.5 ml of serum-free DMEM.
- 4. Wash CEMx174 cells (use 5 x 106 cells for each transfection) twice in serum-free DMEM.
- 5. Add 1.4 ml of the DME/DEAE mix to each 50 µl ligation mix. Vortex gently to mix well.
- 6. Resuspend the cell pellet in the 1.4 ml DNA/DEAE/DMEM mix.
- 7. Incubate for 1 hour at 37°C.
- 8. Centrifuge the cells. Wash once in serum-free DMEM, and once in serum-free RPMI 1640.
- 9. Resuspend the cells in 8-10 ml RPMI 1640 containing 10% fetal bovine serum and pen-strep. Transport the cells to a containment suite, if the procedure was not already performed there.
- 10. Monitor virus growth in the culture every two days (split the cells as needed at the same time). For SHIVs, virus is usually detected after 4-5 days, and will peak in the culture about 7-10 days. SIV is usually a little quicker.

PLASMID DNA

DNA from the plasmids containing the proviral halves can be grown in XL1-Blue bacteria. The bacteria should be grown at 30°C for better yield in DNA preparation.

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